

Keratoacanthoma (KA): An update and review



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Keratoacanthoma (KA) is a common but underreported tumor of the skin. Two striking features of KA are its clinical behavior with spontaneous regression after rapid growth and its nosological position on the border between benignity and malignancy. We review current knowledge on the clinical, histopathological, and dermoscopic features of KA to ensure a proper diagnosis and describe its variants, including different types of multiple KAs. We highlight current concepts of KA ethiopathogenesis with special emphasis on the genetic background of multiple familial KA, the role of Wnt signaling pathway, and induction of KA by BRAF inhibitors and procedures of esthetic dermatology. Finally, treatment strategies are presented with surgical excision as a first option, followed by other modalities, including intralesional chemotherapy, topical and systemic agents, lasers, cryotherapy, and photodynamic therapy. (J Am Acad Dermatol 2016;74:1220-33.)

Key words: BRAF inhibitor; erlotinib; Ferguson-Smith; Grzybowski; histopathology; keratoacanthoma; multiple keratoacanthoma; multiple self-healing squamous epithelioma; squamous cell carcinoma; treatment.

Keratoacanthoma (KA) is common and somewhat cryptic tumor in human beings. Although it had been described already in 1888 by Sir Jonathan Hutchinson,^{1,2} its epidemiology, histopathological diagnostic criteria, prognosis, and treatment guidelines remain controversial.³ Several names used to label KA, including “molluscum sebaceum,” “pseudotumor,” “regressing tumor,” and “self-healing squamous cell carcinoma” (SCC), reflect some of these controversies.^{4,5} The most common concern is related to its position on the border between malignancy and benignity. This imprecision makes it both challenging for a clinician and fascinating for a researcher, as this tumor may hold a key to understanding cancer regression.

EPIDEMIOLOGY

The true incidence of KA is probably underestimated because of misdiagnosis as a SCC, underreporting KA by physicians, or spontaneous regression before the diagnosis can be made. A 2014 study of Carr and Houghton⁶ documented a huge difference in the SCC/KA ratio reported by pathologists from different centers in Great Britain and Ireland. This ratio varied from 2.5:1 to 139:1 and was

Abbreviations used:

GEKA:	generalized eruptive keratoacanthoma
KA:	keratoacanthoma
MSHSE:	multiple self-healing squamous epithelioma
MTS:	Muir-Torre syndrome
SCC:	squamous cell carcinoma

influenced by each pathologist's approach to the diagnosis of KA.

Sporadic solitary KA has an incidence of 104 and 150/100,000 in Australian and Hawaiian populations, respectively.^{7,8} It affects mostly fair-skinned people and was not reported in native Australians. Sun-damaged skin predisposes to KA; its peak incidence has shifted toward the age group 65 to 71 years from 50 to 69 years observed in 1990s.^{3,9} Men are more often affected than women.^{4,9} Variants of KA, including different multiple KA subtypes and KA centrifugum, are rare.

ETIOLOGY

In contrast to ordinary SCC, KA is assumed to originate from the hair follicle.^{3,4} This concept of KA implies it is a benign counterpart of the follicular

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SCC rather than to ordinary SCC.¹⁰ KA exhibits markers that are consistent with those found in the follicular isthmus and infundibulum. Its triphasic nature with proliferative (early, growth), stabilization (well-developed stage), and regression phases has led to the concept of a hair cycle mimicking nature with the KA.^{11,12} In mice with KAs provoked by chemical carcinogens, KA tumor regression was not dependent upon the immune system, but to Wnt/retinoic acid signaling pathways that could support this hypothesis.¹³ The role of the immune system in tumor appearance and resolution is controversial, but has to be considered.^{11,14-17}

Other major signaling pathways potentially involved in KA pathogenesis are summarized in Table I.¹⁸⁻²⁵

Rare cases of solitary and multiple mucous membrane KA, the latter in course of generalized eruptive KA (GEKA) of Grzybowski type, indicate the possibility of different origins of these tumors and differentiation into upper segment hair follicle-like cells.

Genetic background

Predisposing genes for solitary sporadic KA are not known. DNA repair failures associated with the Muir-Torre syndrome (MTS) and xeroderma pigmentosum lead to development of multiple tumors, including KAs.^{3,26,27} Disease-specific mutations in transforming growth factor beta receptor 1 have been described in multiple KAs of Ferguson-Smith type; however, further studies indicated this entity as digenic rather than monogenic.^{28,29}

Provoking factors

Both natural and artificial ultraviolet exposure is a predominant risk factor for KA.^{7,30-34} We have observed development of KA on the x-ray-exposed hand of an interventional radiologist. In fact, roentgen radiation is a well-documented causative factor for KA.^{35,36} Other provoking factors are summarized in Table II.³⁷⁻⁴³

CLINICAL MANIFESTATIONS

Solitary KA

The most common variant of KA is the sporadic and solitary one (Fig 1). Usually it is 1 to 2 cm in

diameter and 0.5 cm in thickness. However, it can vary from a few millimeters up to more than 20 cm in the KA centrifugum variant, also known as KA centrifugum et marginatum.⁴⁴ It cannot be judged at the initial stage how large an individual KA will grow before it will undergo resolution. The first case of KA centrifugum, described by Miedzinski and Kozakiewicz⁴⁴ in Gdańsk, Poland, covered the back of hand. Since then several cases reaching 20 cm in diameter have been documented. The large solitary KA usually has a less regular shape that can mimic a coral reef. If a KA is large and not growing further, one can name them "giant" to distinguish that entity from the constantly enlarging KA centrifugum type.

Rarely solitary KA can be localized on mucous membranes, mostly in the oral cavity, but occasionally on conjunctiva or vulva.⁴⁵⁻⁵⁰

Subungual KAs can be challenging both for diagnosis and management.^{7,51-53}

Multiple KAs

Multiple KAs are rare and can be sporadic or familial. Multiple persistent KAs belong to the first group and are often named "multiple keratoacanthoma centrifugum (et marginatum)" to underline their resemblance to constantly growing neoplasms with a coral reef-like appearance (Fig 2).⁵⁴⁻⁵⁷

Multiple KAs may rarely be associated with prurigo nodularis, usually on the lower limbs of elderly women with sun-damaged skin.⁵⁸

Multiple familial KAs of Ferguson-Smith type, also known as multiple self-healing squamous epithelioma (MSHSE) (Online Mendelian Inheritance in Man [OMIM] 132800), were first described in Scottish families. Genetically confirmed cases have been described worldwide.^{28,59,60} A minor proportion of gene carriers are asymptomatic.

Only around 30 cases of GEKAs of Grzybowski type have been documented (Fig 3).⁶¹⁻⁶³ They were first described in 1950 in Warsaw by Marian Grzybowski.⁶³ Differences between GEKA and MSHSE are presented in Table III. The criteria for the GEKA diagnosis were proposed and divided into mandatory and variable (Table IV).⁶⁴ Visceral

Table I. Cell cycle regulatory pathways involved in the pathogenesis of keratoacanthoma

Pathway	
Wnt	- Wnt is activated in the growth and inactivated in the regression phase. Retinoic acid reverses Wnt-related KA proliferation, promoting tumor regression. That supports the idea of retinoid treatment of KA.
B-Raf	- KAs can appear in course of melanoma treatment with B-Raf inhibitors.
H-ras	- KA has H-ras mutation more commonly than does an ordinary SCC. - HRAS may be involved in the switch from proliferation into regression phase in the biphasic nature of both KA and Spitz nevus. - KAs can appear within nevus sebaceous. Nevus sebaceous is considered to be a segmental RASopathy. These KAs express mutated HRAS.
Hedgehog pathway	- KA can appear in the course of treatment with vismodegib, which is a hedgehog pathway modifier used for the therapy of basal cell carcinoma.
p27	- Cyclin-dependent kinase inhibitor p27 expression is present during the regression stage only, but not during KA growth.

KA, Keratoacanthoma; SCC, squamous cell carcinoma.

Table II. Keratoacanthoma provoking factors

Groups of factors	Provoking factors
Immunosuppression/ immunodeficiency	Iatrogenic: Immunosuppressive and immunomodulatory drugs: Classic immunosuppressant (eg, azathioprine, cyclophosphamide, corticosteroids) Leflunomide Biologic drugs (anti–tumor necrosis factor-alfa) Photochemotherapy Noniatrogenic: Inherited immunodeficiencies Acquired immunodeficiencies: Leprosy, leukemia
Electromagnetic radiation	UVA, UVB, UVC, x-rays including megavoltage radiation
Trauma—koebnerization	Iatrogenic: Surgical procedures Chemical peelings Dermabrasion/microdermabrasion Ablative and coagulating lasers (including fractional lasers) Cryotherapy Photodynamic therapy Irritation after topical drugs (imiquimod) Noniatrogenic: Tattoos Traumas
Chemical factors	Tar
Drugs influencing cell cycle	BRAF inhibitors (vemurafenib, dabrafenib) Hedgehog pathway inhibitor (vismodegib)
Foreign bodies	Tattoos Hyaluronic acid with acrylic hydrogel fillers Collagen fillers

UV, Ultraviolet.

malignancies may be associated with this variant; however, they are far more specific for multiple KAs in the setting of MTS.²⁶

Multiple familial KA of Witten and Zak type is not well characterized. It shares clinical features of

MSHSE and GEKA by having multiple tiny KAs and typical bigger ones coexisting in the same patient. Most cases in the literature can be classified as MSHSE, or as GEKA, or the newly recognized KA associated with prurigo nodularis.^{64–67}



Fig 1. Keratoacanthoma (KA). The solitary and sporadic variant is the most common variant of KA. The lesion starts as a minute papule. The mature KA is a dome- or bud-shaped well-demarcated umbilicated nodule with a hyperkeratotic plug in the center. It is typically localized on sun-exposed areas and evolves in 3 clinical stages: proliferative, mature, and resolving. The process from origin to spontaneous resolution usually occurs within 4 to 6 months and can lead to atrophic hypopigmented scar.

KAs related to predisposing or provoking conditions and factors

Multiple KAs can also appear in the context of rare genetic disorders that predispose to carcinogenesis, such as xeroderma pigmentosum and MTS.^{3,26,27} MTS is characterized by appearance of multiple sebaceous adenomas and other sebaceous tumors along with KAs and high risk of visceral tumors. Subungual KAs were found in patients with familial incontinentia pigmenti.⁶⁸

Iatrogenic KA induced by drugs or medical procedures are summarized in Table II.^{14,20,25,32-34,36,40-42,69-74} With the rapidly growing number of esthetic and antiaging procedures, the risk of inducing KA on sun-damaged skin by laser procedures (mostly resurfacing, including fractional laser), chemical peels, and fillers has to be considered. Koebnerization can also appear in the course of KA treatment with topical drugs (imiquimod) or surgical procedures, increasing the numbers of treatment failures mimicking tumor regrowth after incomplete removal.

DERMOSCOPY

Dermoscopy of KA shares some features with SCC and cannot be used to clearly differentiate these 2 entities.^{75,76} Keratin had the highest sensitivity to differentiate KA and SCC from other amelanotic nodules, and white circles had the highest specificity. Both features together with dot vessels are useful predictors for the diagnosis of KA and SCC. It can help to differentiate other crateriform rapidly growing nodules, including amelanotic melanoma.⁷⁷ Typically, dermoscopy of KA appears as concentric circles of central crater, surrounded by an ivory-whitish area and adjacent peripheral vessels and the most outer circle of whitish halo.⁷⁷

HISTOPATHOLOGICAL EXAMINATION

Diagnosis of the KA is based on 3 principles: typical clinical presentation of a crateriform tumor, rapid (weeks to months) growth with a triphasic course, and histopathological examination of a suitable biopsy specimen.^{78,79} Histopathology of KA is enhanced



Fig 2. Multiple persistent keratoacanthomas (KA). **A**, This 71-year-old woman had multiple KAs varying in size from 1 to 5 cm without a tendency to undergo spontaneous remission and following the typical pattern of lower leg involvement. Some of the KAs aggregated and some lasted for 15 years, with a chronic course of constant growth with some lesions developing a coral-reef appearance as seen in solitary KA centrifugum marginatum. Linked with moderate itching, it was misdiagnosed as prurigo nodularis until the clear morphology of KA became obvious. **B**, Treatment with acitretin 0.5 mg/kg/d for 4 weeks followed by 20 mg/d (0.3 mg/kg/d) for another 8 weeks resulted in marked improvement.



Fig 3. Generalized eruptive keratoacanthoma of Grzybowski. The back of a 51-year-old woman with generalized eruption of hundreds to thousands of small well-demarcated papules, some with keratotic center.

when the clinician is aware of the clinical diagnosis and performs an adequate biopsy.^{4,79} As Dabska and Madejczykowa⁷⁸ emphasized in 1959, the specimen

should be sufficiently representative, including subcutaneous fat, by total or partial excision or by a fusiform partial excision through the entire KA including its center and both sides. This approach allows analysis of not only cellular component, but also its full architecture. Not taking the sample properly may result in a diagnosis of SCC and can lead to overtreatment. Deep shave biopsy of a small KA (during curettage) can be used; however, the deep part of the tumor can be missed.

Cellular characteristics of KA are similar to those of SCC; the architecture is a key feature to establish diagnosis (Fig 4).^{3,80} There are several approaches to distinguish these 2, but none of them have proved to be sufficient.^{16,81-116} The differences between SCC and KA are summarized in Table V.¹¹⁷⁻¹¹⁹

Some dermatopathologists prefer to use the term of “squamous cell carcinoma, keratoacanthoma type” or “probable keratoacanthoma; squamous

Table III. Comparison of multiple familial keratoacanthomas of Ferguson-Smith and generalized eruptive keratoacanthomas of Grzybowski

	KAs of Ferguson-Smith	GEKA of Grzybowski
Family history	Positive, autosomal dominant inheritance	Negative
Onset	First to seventh decade of life	Fifth to seventh decade of life,
Course	KAs slowly grow and resolve within months, but more new KAs are continuously appearing	Eruptive onset and progressive
Lesions:	- Varies among individuals	- Hundreds to thousands
- No.	- Large papules (around 1 cm on the face and bigger on limbs) with horny plug that can fall out leaving an ulcer	- Small, miliary (1-2 mm)
- Size and shape		- Dome-shaped follicular papules with or without keratotic center
Distribution	Extremities and face (nose, ears, circumoral) Trunk is rarely affected and palms and soles are not affected	Generalized; sun-exposed areas including upper aspect of trunk and face; intertriginous areas
Scarring	Pitted scars, more disfiguring on the face than on the limbs	Not pronounced
Pruritus	Not present	Prominent

GEKA, Generalized eruptive keratoacanthoma; KA, keratoacanthoma.

Table IV. Diagnostic criteria for multiple eruptive keratoacanthomas proposed by Nofal et al⁶⁴ and modified by the sixth consistent criteria: lack of family history of multiple keratoacanthomas

Consistent criteria
Onset in adulthood (usually fifth to seventh decade of life)
Generalized eruption of hundreds to thousands of small well-demarcated papules, some with a keratotic center
Progressive course
Severe and persistent pruritus
Histopathology consistent with KA
Lack of family history of multiple KA
Variable criteria
Masked face ("mask of Zorro" sign)*
Mucosal lesions
Crateriform nodules (typical solitary KA)
Ectropion

KA, Keratoacanthoma.

*Included into variable criteria is extensive facial involvement that affects predominantly periorbital region.

cell carcinoma cannot be ruled out." The study of Carr and Houghton⁶ placed emphasis on the importance of clinical description, quality of the specimen obtained with excisional biopsy, and evidence of regression favoring KA diagnosis. The last feature decreases the ratio of SCC/KA diagnosis in those centers with longer wait times for surgery.

A variety of different immunohistochemical staining for several markers has been used to help to distinguish KA from SCC (Table VI).^{16,90,93,101,103,106,108,109,112,113,116,120-127} Unfortunately, the huge number of potentially specific markers used is testimony to the fact that there is no really adequate one.

Histopathology is necessary to rule out other diseases that can present as crateriform papules or nodules (Table VII),^{53,118,128-140} and can delineate variants of SCC, especially the follicular SCC, which may mimic KA.^{139,140}

KA TREATMENT

Solitary KA

Controversies remain about the management of the solitary KA. A wait-and-see strategy for a solitary KA, which assumes spontaneous regression, is questionable unless clear signs of involution are already present.¹⁴¹ One cannot predict the final size of a KA that can reach several centimeters before it will regress, leaving a potentially disfiguring scar. The potential for transformation into invasive SCC with metastases is extremely low; however, it has to be considered when choosing the treatment.¹⁴²⁻¹⁴⁴ Interestingly, there has been no single case of KA reported in the literature that has led to fatal outcome, as reviewed by Savage and Maize¹⁴⁵ in 2014. It has to be mentioned that the authors did not include the patient who died from metastatic SCC described by Hodak et al¹⁴⁴ because of an inadequately proven link between KA and metastases. The diagnosis of KA should correlate clinical and histologic findings, implying a tissue specimen is highly desirable before or concurrent with treatment.

Whenever possible, surgical treatment is a gold standard regimen with full-thickness fusiform excision providing good esthetic outcome and an optimal specimen for the pathologist. Unfortunately, there are no specific margins established for KA, but the same as for noninvasive SCC can be advised (5 mm) to assure 95% chance of complete

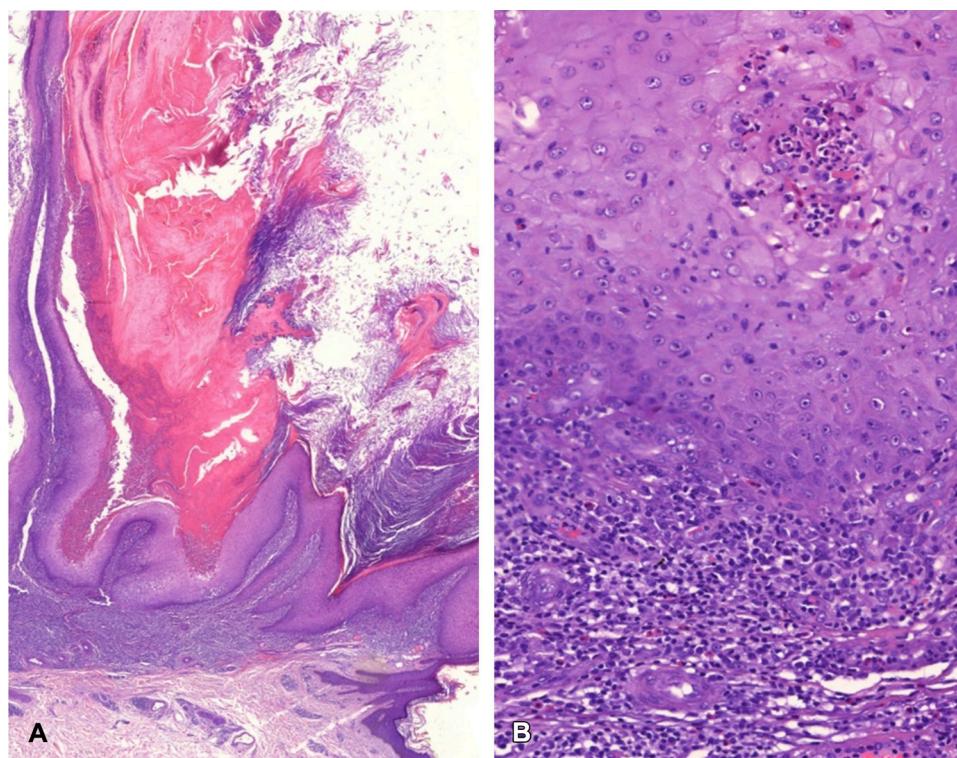


Fig 4. Histopathology of keratoacanthoma (KA). **A**, Epithelial lip at the periphery that extends partially over the central keratin plug and dense lichenoid infiltrate. **B**, Glassy appearance of keratinocytes along with lymphocytic and eosinophilic infiltrate. Intraepithelial microabscess is not always judged as a hallmark of KA.

Table V. Histopathological features used for keratoacanthoma and squamous cell carcinoma differentiation

Histologic feature	KA vs SCC
Symmetry*	KA>>SCC
Epithelial lipping*	KA>FSCC>SCC
Sharp demarcation between tumor and stroma*	KA>SCC
Ulceration*	KA<SCC
Mitoses*	KA<SCC
Pleomorphism/anaplasia*	KA<SCC (SCC randomly scattered; KA gradually increased in deeper parts of tumor)
Ground-glass appearance*	KA>FSCC>SCC
Intraepithelial elastic fibers	KA>SCC
Keratin-filled crater	KA>SCC
Extension beyond sweat glands	KA<SCC
Intraepithelial abscess	KA=SCC or KA>SCC depending on author
Lateral growth predominance	KA=SCC
Dyskeratosis	KA=SCC, KA<FSCC
Parakeratosis	KA=SCC or KA<SCC depending on author
Acantholysis	KA<SCC; FSCC has intraepithelial mucin in association with acantholysis in >50% of cases, which is not seen in KA
Perineural invasion	SCC, aggressive behavior
	KA, not consistent sign of aggressive behavior

FSCC, Follicular squamous cell carcinoma; KA, keratoacanthoma; SCC, squamous cell carcinoma.

*None of the 16 histologic features can be used as a clear-cut criterion for the diagnosis or exclusion of keratoacanthoma, but the most valuable ones are marked with asterisks. FSCC can clinically mimic keratoacanthoma in 15% of cases. The most distinctive features of this variant important for differential diagnosis of keratoacanthoma are indicated.

Table VI. Spectrum of selected histochemical and cytogenetic markers that were proposed to be of value in keratoacanthoma/squamous cell carcinoma differentiation

Group of markers	Marker	KA/SCC
Inflammatory infiltrate		
Plasmacytoid dendritic cells	BDCA2	KA>SCC
Regulatory T cells	CD3 ⁺ FoxP3 ⁺	KA<SCC
Macrophages	CD163	KA<SCC
IL-27-producing cells	IL-27	KA>SCC
Apoptosis/cell death		
Bcl-2	KA<SCC	
Bcl-x	KA<SCC	
AIF	KA<SCC	
TUNEL	KA>SCC	
Le ^y	KA>SCC	
Cyclin A and B	KA>SCC	
P2X ₇	KA>SCC	
Proliferation		
NUCKS	KA<SCC	
PCNA	Different patterns	
IMP3	KA<SCC	
Differentiation		
Keratins	Outer root sheath beneath infundibulum pattern of KA and epidermis pattern of SCC	
Filaggrin	KA>SCC	
Signaling		
p53	KA<SCC	
p50	KA>SCC	
Adhesion and migration		
VCAM (CD106)	Expression associated with triphasic nature of KA and maturation status of SCC	
ICAM (CD54)	KA>invasive SCC	
Syndecan-1	Different pattern	
Desmoglein 1 and 2	KA>SCC	
E-cadherin	KA>SCC	
Cortactin	KA>SCC	
Lectins	KA>SCC	
Genetic		
Microsatellite instability and loss of heterogeneity	KA in the setting of Muir-Torre syndrome	
Other		
Oncostatin M on:	KA>SCC	
- Tumor cells	KA<SCC	
- Macrophages	KA<SCC	
- COX2		

AIF, Apoptosis inducing factor; *Bcl-2*, B-cell chronic lymphocytic leukemia/lymphoma-2; *Bcl-x*, B-cell chronic lymphocytic leukemia/lymphoma-2 related genes; *BDCA*, blood dendritic cell antigen; *COX-2*, cyclooxygenase 2; *FoxP3*, fork-head box P3; *ICAM*, intercellular adhesion molecule; *IMP3*, insulin-like growth factor II mRNA binding protein 3; *IL*, interleukin; *KA*, keratoacanthoma; *Le^y*, Lewis- y; *NUCS*, nuclear ubiquitous casein and cyclin-dependent kinases substrate; *P2X₇*, ionotropic P2 receptor family 7; *PCNA*, proliferating cell nuclear antigen; *SCC*, squamous cell carcinoma; *TUNEL*, terminal uridine nick-end labeling; *VCAM*, vascular cell adhesion molecule.

removal.¹⁴⁶⁻¹⁴⁸ Negative margins are of predictive value for complete removal. Positive margins usually do not indicate tumor recurrence.¹⁴⁹ Mohs micrographic technique is desirable for large KA (including KA centrifugum) and/or those in cosmetically sensitive areas.¹⁵⁰ Deep curettage of an entire KA can be an alternative approach for small

ones, but has to be followed by histologic evaluation. Paradoxically, curettage is probably increasing the ratio of SCC:KA in pathology reports, as dermatopathologists tend to overdiagnose SCC when they do not have a full-thickness specimen.⁶

We often use intralesional chemotherapy after obtaining a properly performed incisional skin

Table VII. Differential diagnosis of keratoacanthoma

Crateriform lesions with SCC features	KA-like SCC KA with malignant transformation Follicular (infundibular) SCC (crateriform) Crateriform SCC arisen from actinic keratosis Crateriform Bowen disease Verrucous carcinoma Onycholemmal carcinoma (for subungual KA)
Other tumors	Exophytic pilomatrixoma Cutaneous metastatic disease Amelanotic melanoma Primary cutaneous CD30 anaplastic large-cell lymphoma
Infectious diseases	Cryptococcosis Chromoblastomycosis Sporotrichosis North American blastomycosis Tuberculosis verrucosa cutis Giant molluscum contagiosum
Inflammatory diseases	Prurigo nodularis Hypertrrophic discoid lupus erythematosus Hypertrrophic lichen planus Halogenoderma

KA, Keratoacanthoma; SCC, squamous cell carcinoma.

biopsy specimen, following the regimen originated by Klein et al.¹⁵¹ It is the second-line option of KA treatment, but evidence of efficacy is limited.¹⁵² Methotrexate and 5-fluorouracil are preferred as intralesional drugs, with bleomycin or interferons being another option. Methotrexate usually requires 2 or more injections to obtain remission. Intralesional chemotherapy can precede surgery to reduce the size of tumor of about 50% to 80% before the excision. Two-step regimen provides a better cosmetic and functional outcome than intralesional treatment alone.¹⁵⁵ Other therapeutic modalities, including those preferred for KA centrifugum, are summarized in Table VIII.^{74,151,156-166}

Multiple KAs

Systemic acitretin or other retinoids are a first-line option for variants of multiple KA, as monotherapy or combined with surgery or other second-line procedures as for solitary tumor.^{167,168} The dosage varies from 0.5 to 1.0 mg/kg of acitretin at the beginning of treatment and can be tapered as needed. A marked response is usually evident; however, total long-lasting clearance is hardly ever achieved (Fig 2, B). Smaller doses of 10 to 20 mg/d of acitretin or repeated courses of treatment are often

necessary to sustain clinical response. Resistant KAs may occur, especially in GEKA of Grzybowski,¹⁶⁹ and require other approaches.

Reports on the use of systemic cytostatic agents, such as systemic methotrexate and 5-fluorouracil, are anecdotal. In contrast to intralesional methotrexate, the efficacy of systemic methotrexate is less predictable. Cyclophosphamide was shown to be effective in retinoid- and methotrexate-resistant cases of multiple KAs with pulses of 1 g per month given to reduce cumulative dose and the risk of long-term toxicity.¹⁶⁹ Erlotinib, an epidermal growth factor receptor inhibitor, is a new promising approach for resistant KAs; however, experience with it is still limited.¹⁷⁰ Intralesional corticosteroids are occasionally used in GEKA of Grzybowski type with good response either as monotherapy or with systemic retinoids. The latter combined treatment is a good option in KA arising in the setting of prurigo nodularis.^{58,171} The addition of cyclosporine to systemic treatment in these cases may also be considered.⁵⁸

KAs in patients treated with BRAF inhibitor

Solitary KAs related to BRAF inhibitor therapy were successfully treated with total surgical excision and photodynamic therapy. Multiple KAs can be also handled with systemic retinoids combined with intralesional 5-fluorouracil.^{128,172} The appearance of KAs in this setting should not influence melanoma treatment.

Follow-up

Patients should be monitored after KA removal. The recurrence rate ranges from 1% to 8%. In addition, a new KA can appear at the site of treatment within 1 week and 8 months because of koebnerization sometimes evident after surgery, cryotherapy, imiquimod, and photodynamic therapy.¹⁷³ Patients should be advised to avoid provoking factors, including intense and prolonged ultraviolet light exposure, and to perform self-evaluation in all predisposed areas. Patients with the history of KA should be informed about the higher risk of new KA appearance after traumatizing medical or cosmetic procedures performed on photo-damaged skin.

Conclusion

Rare cases of KA that evolved into SCC or that behaved as malignant tumors have changed the clinical perspective of KA during the last 30 years.^{142-144,174} The management of KA has evolved toward that used for well-differentiated SCC.¹⁷⁵ This approach is suitable as long as it does not compromise functional and esthetic outcome more than should be expected during the natural

Table VIII. Alternative to surgery and intralesional chemotherapeutics treatments for solitary keratoacanthoma

Treatment modality	
Ablative lasers	Both modalities are suitable for small KAs when surgery is not available or possible and should be preceded with histopathological examination
Cryotherapy	Postsurgery when aggressive course is predicted or other options are contraindicated
Radiotherapy	Several sessions are required to obtain remission
Photodynamic therapy	Cases of aggravation or induction of KA by this regimen have been described
Topical treatments:	Can be used as monotherapy or sequential after ablative laser or other destructive techniques
- 5-Fluorouracil	For imiquimod, 4-11 wk of application on a daily basis or every 2-3 d is necessary to obtain complete remission
- Imiquimod	
- Podophyllin	
Systemic erlotinib	Epidermal growth factor receptor inhibitor can be used for KA centrifugum when an aggressive course is evident and surgery or combined intralesional treatment and surgery are not possible or are ineffective
Systemic retinoids	In KA centrifugum when other options are not available or contraindicated

KA, Keratoacanthoma.

course of the KA. KA should be regarded as a separate entity with a distinct clinical appearance and course. Many studies addressing the problem of clear histopathological differentiation between SCC and KA support the concept of the peculiarity and importance of KA as a precise diagnosis. In KA, diagnosis should be based on clinical and pathological correlation. A correct diagnosis of KA discourages overtreatment, and provides a wider spectrum of treatment approaches than those recommended for various types of SCC.

REFERENCES

- Hutchinson J. Morbid growths and tumors: 1: the "crateriform ulcer of the face" a form of acute epithelial cancer. *Trans Pathol Soc London.* 1889;40:275-281.
- Hutchinson J. *A peculiar form of cancer of the skin: illustrations of clinical surgery consisting of plates, photographs, woodcuts, diagrams, etc. illustrating surgical diseases, symptoms and accidents also operative and other methods of treatment with descriptive letterpress.* Vol 1. Philadelphia (PA): P Blakiston and Son; 1888. Plate 92.
- Schwartz RA. Keratoacanthoma: a clinico-pathologic enigma. *Dermatol Surg.* 2004;30:326-333.
- Schwartz RA. Keratoacanthoma. *J Am Acad Dermatol.* 1994; 30:1-19. quiz 20-22.
- Mandrell JC, Santa Cruz D. Keratoacanthoma: hyperplasia, benign neoplasm, or a type of squamous cell carcinoma? *Semin Diagn Pathol.* 2009;26:150-163.
- Carr RA, Houghton JP. Histopathologists' approach to keratoacanthoma: a multisite survey of regional variation in Great Britain and Ireland. *J Clin Pathol.* 2014;67:637-638.
- Sullivan JJ. Keratoacanthoma: the Australian experience. *Australas J Dermatol.* 1997;38(Suppl 1):S36-S39.
- Reizner GT, Chuang TY, Elpern DJ, Stone JL, Farmer ER. Basal cell carcinoma and keratoacanthoma in Hawaiians: an incidence report. *J Am Acad Dermatol.* 1993;29:780-782.
- Vergilis-Kalner IJ, Kriseman Y, Goldberg LH. Keratoacanthomas: overview and comparison between Houston and Minneapolis experiences. *J Drugs Dermatol.* 2010;9:117-121.
- Misago N, Inoue T, Nagase K, et al. Crater/ulcerated form of infundibular squamous cell carcinoma: a possible distinct entity as a malignant (or high-grade) counterpart to keratoacanthoma. *J Dermatol.* 2015;42:667-673.
- Ramselaar CG, Ruitenberg EJ, Kruizinga W. Regression of induced keratoacanthomas in anagen (hair growth phase) skin grafts in mice. *Cancer Res.* 1980;40:1668-1673.
- Iyengar B, Ramesh V. Hair cycle and the histogenesis of pillar tumors. *Indian J Cancer.* 1989;26:1-9.
- Zito G, Saotome I, Liu Z, et al. Spontaneous tumor regression in keratoacanthomas is driven by Wnt/retinoic acid signaling cross-talk. *Nat Commun.* 2014;5:3543.
- Lain EL, Markus RF. Early and explosive development of nodular basal cell carcinoma and multiple keratoacanthomas in psoriasis patients treated with cyclosporine. *J Drugs Dermatol.* 2004;3:680-682.
- Walder BK, Robertson MR, Jeremy D. Skin cancer and immunosuppression. *Lancet.* 1971;2:1282-1283.
- Kambayashi Y, Fujimura T, Aiba S. Comparison of immunosuppressive and immunomodulatory cells in keratoacanthoma and cutaneous squamous cell carcinoma. *Acta Derm Venereol.* 2013;93:663-668.
- Prehn RT. The paradoxical association of regression with a poor prognosis in melanoma contrasted with a good prognosis in keratoacanthoma. *Cancer Res.* 1996;56:937-940.
- Hu W, Cook T, Oh CW, Penneys NS. Expression of the cyclin-dependent kinase inhibitor p27 in keratoacanthoma. *J Am Acad Dermatol.* 2000;42:473-475.
- Corominas M, Kamino H, Leon J, Pellicer A. Oncogene activation in human benign tumors of the skin (keratoacanthomas): is HRAS involved in differentiation as well as proliferation? *Proc Natl Acad Sci U S A.* 1989;86:6372-6376.
- Aasi S, Silkiss R, Tang JY, et al. New onset of keratoacanthomas after vismodegib treatment for locally advanced basal cell carcinomas: a report of 2 cases. *JAMA Dermatol.* 2013;149: 242-243.
- Su F, Viros A, Milagre C, et al. RAS mutations in cutaneous squamous-cell carcinomas in patients treated with BRAF inhibitors. *N Engl J Med.* 2012;366:207-215.
- Zalaudek I, Bonifazi E, Ferrara G, Argenziano G. Keratoacanthomas and Spitz tumors: are they both 'self-limiting' variants of malignant cutaneous neoplasms? *Dermatology.* 2009;219:3-6.

23. Cribier B, Scrivener Y, Grosshans E. Tumors arising in nevus sebaceus: a study of 596 cases. *J Am Acad Dermatol.* 2000;42: 263-268.
24. Long GV, Trefzer U, Davies MA, et al. Dabrafenib in patients with Val600Glu or Val600Lys BRAF-mutant melanoma metastatic to the brain (BREAK-MB): a multicenter, open-label, phase 2 trial. *Lancet Oncol.* 2012;13:1087-1095.
25. Chapman PB, Hauschild A, Robert C, et al. Improved survival with vemurafenib in melanoma with BRAF V600E mutation. *N Engl J Med.* 2011;364:2507-2516.
26. Schwartz RA, Torre DP. The Muir-Torre syndrome: a 25-year retrospect. *J Am Acad Dermatol.* 1995;33:90-104.
27. Schwartz RA. The keratoacanthoma: a review. *J Surg Oncol.* 1979;12:305-317.
28. Goudie DR, D'Alessandro M, Merriman B, et al. Multiple self-healing squamous epithelioma is caused by a disease-specific spectrum of mutations in TGFBR1. *Nat Genet.* 2011;43:365-369.
29. Kang HC, Quigley DA, Kim IJ, et al. Multiple self-healing squamous epithelioma (MSSE): rare variants in an adjacent region of chromosome 9q22.3 to known TGFBR1 mutations suggest a digenic or multilocus etiology. *J Invest Dermatol.* 2013;133:1907-1910.
30. Dufresne RG, Marrero GM, Robinson-Bostom L. Seasonal presentation of keratoacanthomas in Rhode Island. *Br J Dermatol.* 1997;136:227-229.
31. Wolfe CM, Green WH, Cognetta AB Jr, Hatfield HK. Multiple squamous cell carcinomas and eruptive keratoacanthomas in an arc welder. *Dermatol Surg.* 2013;39:328-330.
32. Sina B, Adrian RM. Multiple keratoacanthomas possibly induced by psoralens and ultraviolet A photochemotherapy. *J Am Acad Dermatol.* 1983;9:686-688.
33. Craddock KJ, Rao J, Lauzon GJ, Tron VA. Multiple keratoacanthomas arising post-UVB therapy. *J Cutan Med Surg.* 2004; 8:239-243.
34. Brazzelli V, Barbagallo T, Prestinari F, et al. Keratoacanthoma in vitiligo lesion after UVB narrowband phototherapy. *Photodermat Photoimmunol Photomed.* 2006;22:211-213.
35. Robertson SJ, Bashir SJ, Pichert G, Robson A, Whittaker S. Severe exacerbation of multiple self-healing squamous epithelioma (Ferguson-Smith disease) with radiotherapy, which was successfully treated with acitretin. *Clin Exp Dermatol.* 2010;35:e100-e102.
36. Shaw JC, Storrs FJ, Everts E. Multiple keratoacanthomas after megavoltage radiation therapy. *J Am Acad Dermatol.* 1990;23: 1009-1011.
37. Pattee SF, Silvis NG. Keratoacanthoma developing in sites of previous trauma: a report of two cases and review of the literature. *J Am Acad Dermatol.* 2003;48(Suppl):S35-S38.
38. Mohr B III, Fernandez MP, Krejci-Manwaring J. Eruptive keratoacanthomas after Jessners and trichloroacetic acid peel for actinic keratosis. *Dermatol Surg.* 2013;39:331-333.
39. Cox S. Rapid development of keratoacanthomas after a body peel. *Dermatol Surg.* 2003;29:201-203.
40. Mamelak AJ, Goldberg LH, Marquez D, Hosler GA, Hinckley MR, Friedman PM. Eruptive keratoacanthomas on the legs after fractional photothermolysis: report of two cases. *Dermatol Surg.* 2009;35:513-518.
41. Goldberg LH, Silapunt S, Beyrau KK, Peterson SR, Friedman PM, Alam M. Keratoacanthoma as a postoperative complication of skin cancer excision. *J Am Acad Dermatol.* 2004;50:753-758.
42. Gewirtzman A, Meirson DH, Rabinovitz H. Eruptive keratoacanthomas following carbon dioxide laser resurfacing. *Dermatol Surg.* 1999;25:666-668.
43. Ghadially FN, Barton BW, Kerridge DF. The etiology of keratoacanthoma. *Cancer.* 1963;16:603-611.
44. Miedzinski F, Kozakiewicz J. Keratoacanthoma centrifugum—a special variety of keratoacanthoma [in German]. *Hautarzt.* 1962;13:348-352.
45. Ozkan F, Bilgic R, Cesur S. Vulvar keratoacanthoma. *APMIS.* 2006;114:562-565.
46. D'Agostino G, Colonna M. Keratoacanthoma of the vulva [in Italian]. *Gazz Int Med Chir.* 1962;67:1032-1038.
47. Svirsky JA, Freedman PD, Lumerman H. Solitary intraoral keratoacanthoma. *Oral Surg Oral Med Oral Pathol.* 1977;43: 116-122.
48. Hamed LM, Wilson FM II, Grayson M. Keratoacanthoma of the limbus. *Ophthalmic Surg.* 1988;19:267-270.
49. Eversole LR, Leider AS, Alexander G. Intraoral and labial keratoacanthoma. *Oral Surg Oral Med Oral Pathol.* 1982;54: 663-667.
50. Drizhal I, Nozicka Z. Keratoacanthoma of mucous membrane. A case report. *Sb Ved Pr Lek Fak Karlovy Univerzity Hradci Kralove.* 1973;16:151-154.
51. Perrin C. Tumors of the nail unit. A review. Part II: acquired localized longitudinal pachyonychia and masked nail tumors. *Am J Dermatopathol.* 2013;35:693-709; quiz 10-12.
52. Lamp JC, Graham JH, Urbach F, Burgoon CF Jr. Keratoacanthoma of the subungual region. A clinicopathological and therapeutic study. *J Bone Joint Surg Am.* 1964;46:1721-1731.
53. Chaser BE, Renszel KM, Crowson AN, et al. Onycholemmal carcinoma: a morphologic comparison of 6 reported cases. *J Am Acad Dermatol.* 2013;68:290-295.
54. Schwartz RA. Multiple persistent keratoacanthomas. *Oncology.* 1979;36:281-285.
55. Divers AK, Correale D, Lee JB. Keratoacanthoma centrifugum marginatum: a diagnostic and therapeutic challenge. *Cutis.* 2004;73:257-262.
56. Khaled A, Kourda M, Fazaa B, Zermani R, Kamoun MR. Multiple keratoacanthoma centrifugum marginatum. *Dermatol Online J.* 2010;16:16.
57. Ogasawara Y, Kinoshita E, Ishida T, Hamamoto Y, Fujiyama J, Muto M. A case of multiple keratoacanthoma centrifugum marginatum: response to oral etretinate. *J Am Acad Dermatol.* 2003;48:282-285.
58. Wu TP, Miller K, Cohen DE, Stein JA. Keratoacanthomas arising in association with prurigo nodules in pruritic, actinally damaged skin. *J Am Acad Dermatol.* 2013;69:426-430.
59. Ferguson Smith J. A case of multiple primary squamous-celled carcinomata of the skin in a young man, with spontaneous healing. *Br J Dermatol Syphil.* 1934;46:267-272.
60. Ferguson-Smith MA, Wallace DC, James ZH, Renwick JH. Multiple self-healing squamous epithelioma. *Birth Defects Orig Artic Ser.* 1971;7:157-163.
61. Anzalone CL, Cohen PR. Generalized eruptive keratoacanthomas of Grzybowski. *Int J Dermatol.* 2014;53:131-136.
62. Schwartz RA, Blaszczyk M, Jablonska S. Generalized eruptive keratoacanthoma of Grzybowski: follow-up of the original description and 50-year retrospect. *Dermatology.* 2002;205: 348-352.
63. Grzybowski M. A case of peculiar generalized epithelial tumors of the skin. *Br J Dermatol Syphil.* 1950;62:310-313.
64. Nofal A, Assaf M, Nofal E, Alradi M. Generalized eruptive keratoacanthoma: proposed diagnostic criteria and therapeutic evaluation. *J Eur Acad Dermatol Venereol.* 2014;28: 397-404.
65. Agarwal M, Chander R, Karmakar S, Walia R. Multiple familial keratoacanthoma of Witten and Zak - a report of three siblings. *Dermatology.* 1999;198:396-399.

66. Witten VH, Zak FG. Multiple, primary, self-healing prickle-cell epithelioma of the skin. *Cancer*. 1952;5:539-550.
67. Boateng B, Hornstein OP, von den Driesch P, Kiesewetter F. Multiple keratoacanthomas (Witten-Zak type) in prurigo simplex subacuta [in German]. *Hautarzt*. 1995;46:114-117.
68. Montes CM, Maize JC, Guerry-Force ML. Incontinentia pigmenti with painful subungual tumors: a two-generation study. *J Am Acad Dermatol*. 2004;50(Suppl):S45-S52.
69. Esser AC, Abril A, Fayne S, Doyle JA. Acute development of multiple keratoacanthomas and squamous cell carcinomas after treatment with infliximab. *J Am Acad Dermatol*. 2004;50(Suppl):S75-S77.
70. Anforth RM, Blumetti TC, Kefferd RF, et al. Cutaneous manifestations of dabrafenib (GSK2118436): a selective inhibitor of mutant BRAF in patients with metastatic melanoma. *Br J Dermatol*. 2012;167:1153-1160.
71. Gamo R, Pinedo F, Vicente J, et al. Keratoacanthoma-like reaction after a hyaluronic acid and acrylic hydrogel cosmetic filler. *Dermatol Surg*. 2008;34:954-959.
72. Campalani E, Holden CA. Keratoacanthoma associated with the use of topical imiquimod. *Clin Exp Dermatol*. 2013;38:555-556.
73. Pini AM, Koch S, Scherer L, French LE, Lauchli S, Hofbauer GF. Eruptive keratoacanthoma following topical imiquimod for *in situ* squamous cell carcinoma of the skin in a renal transplant recipient. *J Am Acad Dermatol*. 2008;59(Suppl):S116-S117.
74. Gogia R, Grekin RC, Shinkai K. Eruptive self-resolving keratoacanthomas developing after treatment with photodynamic therapy and microdermabrasion. *Dermatol Surg*. 2013;39:1717-1720.
75. Zalaudek I, Giacomel J, Schmid K, et al. Dermatoscopy of facial actinic keratosis, intraepidermal carcinoma, and invasive squamous cell carcinoma: a progression model. *J Am Acad Dermatol*. 2012;66:589-597.
76. Rosendahl C, Cameron A, Argenziano G, Zalaudek I, Tschandl P, Kittler H. Dermoscopy of squamous cell carcinoma and keratoacanthoma. *Arch Dermatol*. 2012;148:1386-1392.
77. Cavicchini S, Tourlaki A, Lunardon L, Boneschi V, Gianotti R. Amelanotic melanoma mimicking keratoacanthoma: the diagnostic role of dermoscopy. *Int J Dermatol*. 2013;52:1023-1024.
78. Dabska M, Madejczykowa A. Rogowiak kolczastokomorkowy: keratoacanthoma (molluscum sebaceum, molluscum pseudocarcinomatous). Studium patologiczno-kliniczne. *Nowotwory*. 1959;9:1-23.
79. Dabska M. Keratoacanthoma [in Polish]. *Wiad Lek*. 1965;18:1249-1250.
80. Kern WH, McCray MK. The histopathologic differentiation of keratoacanthoma and squamous cell carcinoma of the skin. *J Cutan Pathol*. 1980;7:318-325.
81. Sagol O, Kurtoglu B, Ozer E, Pabuccuoglu U. Stereological estimation of mean nuclear volume and staining pattern of Ki-67 antigen in keratoacanthomas and squamous cell carcinomas. *Gen Diagn Pathol*. 1998;143:305-309.
82. Phillips P, Helm KF. Proliferating cell nuclear antigen distribution in keratoacanthoma and squamous cell carcinoma. *J Cutan Pathol*. 1993;20:424-428.
83. Ellis GL. Differentiating keratoacanthoma from squamous cell carcinoma of the lower lip: an analysis of intraepithelial elastic fibers and intracytoplasmic glycogen. *Oral Surg Oral Med Oral Pathol*. 1983;56:527-532.
84. Binder M, Steiner A, Mossbacher U, Hunegnaw M, Pehamberger H, Wolff K. Estimation of the volume-weighted mean nuclear volume discriminates keratoacanthoma from squamous cell carcinoma. *Am J Dermatopathol*. 1998;20:453-458.
85. Aung PP, Ballester LY, Mahalingam M. Histopathology of keratoacanthoma revisited: utility of orificial size as a diagnostic adjunct. *Int J Surg Pathol*. 2013;22:316-325.
86. Kwittken J. A histologic chronology of the clinical course of the keratocarcinoma (so-called keratoacanthoma). *M Sinai J Med*. 1975;42:127-135.
87. Milewski B, Chorzelski T. Comparative histological and histochemical studies on keratoacanthoma and highly differentiated spinocellular epithelioma [in German]. *Hautarzt*. 1962;13:7-12.
88. Cribier B, Asch P, Grosshans E. Differentiating squamous cell carcinoma from keratoacanthoma using histopathological criteria. Is it possible? A study of 296 cases. *Dermatology*. 1999;199:208-212.
89. Asch PH, Basset P, Roos M, Grosshans E, Bellocq JP, Cribier B. Expression of stromelysin 3 in keratoacanthoma and squamous cell carcinoma. *Am J Dermatopathol*. 1999;21:146-150.
90. Kannon G, Park HK. Utility of peanut agglutinin (PNA) in the diagnosis of squamous cell carcinoma and keratoacanthoma. *Am J Dermatopathol*. 1990;12:31-36.
91. Weedon DD, Malo J, Brooks D, Williamson R. Squamous cell carcinoma arising in keratoacanthoma: a neglected phenomenon in the elderly. *Am J Dermatopathol*. 2010;32:423-426.
92. Sanchez Yus E, Simon P, Requena L, Ambrojo P, de Eusebio E. Solitary keratoacanthoma: a self-healing proliferation that frequently becomes malignant. *Am J Dermatopathol*. 2000;22:305-310.
93. Tsuji T. Keratoacanthoma and squamous cell carcinoma: study of PCNA and Le(Y) expression. *J Cutan Pathol*. 1997;24:409-415.
94. Peris K, Magrini F, Keller G, et al. Analysis of microsatellite instability and loss of heterozygosity in keratoacanthoma. *Arch Dermatol Res*. 1997;289:185-188.
95. Yao D, Alexander CL, Quinn JA, Chan WC, Wu H, Greenhalgh DA. Fos cooperation with PTEN loss elicits keratoacanthoma not carcinoma, owing to p53/p21 WAF-induced differentiation triggered by GSK3beta inactivation and reduced AKT activity. *J Cell Sci*. 2008;121:1758-1769.
96. Vasiljevic N, Andersson K, Bjelkenkrantz K, et al. The Bcl-xL inhibitor of apoptosis is preferentially expressed in cutaneous squamous cell carcinoma compared with that in keratoacanthoma. *Int J Cancer*. 2009;124:2361-2366.
97. Stephenson TJ, Royds JA, Bleehen SS, Silcocks PB, Rees RC. 'Anti-metastatic' nm23 gene product expression in keratoacanthoma and squamous cell carcinoma. *Dermatology*. 1993;187:95-99.
98. Stephenson TJ, Royds J, Silcocks PB, Bleehen SS. Mutant p53 oncogene expression in keratoacanthoma and squamous cell carcinoma. *Br J Dermatol*. 1992;127:566-570.
99. Putti TC, Teh M, Lee YS. Biological behavior of keratoacanthoma and squamous cell carcinoma: telomerase activity and COX-2 as potential markers. *Mod Pathol*. 2004;17:468-475.
100. Lowry WS, Atkinson RJ. Mutant p53 oncogene expression in keratoacanthoma and squamous cell carcinoma. *Br J Dermatol*. 1993;128:708.
101. Lee YS, Teh M. p53 Expression in pseudoepitheliomatous hyperplasia, keratoacanthoma, and squamous cell carcinoma of skin. *Cancer*. 1994;73:2317-2323.
102. LeBoit PE. Is keratoacanthoma a variant of squamous cell carcinoma. New insights into an old controversy... soon? *Am J Dermatopathol*. 1995;17:319-320.

103. Kerschmann RL, McCalmont TH, LeBoit PE. p53 Oncoprotein expression and proliferation index in keratoacanthoma and squamous cell carcinoma. *Arch Dermatol.* 1994;130:181-186.
104. Connolly M, Narayan S, Oxley J, de Berker DA. Immunohistochemical staining for the differentiation of subungual keratoacanthoma from subungual squamous cell carcinoma. *Clin Exp Dermatol.* 2008;33:625-628.
105. Cain CT, Niemann TH, Argenyi ZB. Keratoacanthoma versus squamous cell carcinoma. An immunohistochemical reappraisal of p53 protein and proliferating cell nuclear antigen expression in keratoacanthoma-like tumors. *Am J Dermatopathol.* 1995;17:324-331.
106. Batinac T, Zamolo G, Coklo M, Hadzisejdic I, Stemberger C, Zauhar G. Expression of cell cycle and apoptosis regulatory proteins in keratoacanthoma and squamous cell carcinoma. *Pathol Res Pract.* 2006;202:599-607.
107. Cabibbi D, Aragona F, Guarinotta C, et al. Glut-1 expression and in situ CD1a/CD57 immunologic deficit in keratoacanthoma and squamous cell carcinoma of immunocompetent patients. *Appl Immunohistochem Mol Morphol.* 2011;19:239-245.
108. Fujii M, Honma M, Takahashi H, Ishida-Yamamoto A, Iizuka H. The nuclear factor kappa B p50 subunit and cortactin as markers to distinguish between keratoacanthoma and well-differentiated squamous cell carcinoma. *Clin Exp Dermatol.* 2011;36:788-792.
109. Cabrijan L, Lipozencic J, Batinac T, Lenkovic M, Zgombic ZS. Differences between keratoacanthoma and squamous cell carcinoma using TGF-alpha. *Coll Antropol.* 2013;37:147-150.
110. Waring AJ, Takata M, Rehman I, Rees JL. Loss of heterozygosity analysis of keratoacanthoma reveals multiple differences from cutaneous squamous cell carcinoma. *Br J Cancer.* 1996;73:649-653.
111. Biesterfeld S, Josef J. Differential diagnosis of keratoacanthoma and squamous cell carcinoma of the epidermis by MIB-1 immunohistometry. *Anticancer Res.* 2002;22:3019-3023.
112. Patel A, Halliday GM, Cooke BE, Barnetson RS. Evidence that regression in keratoacanthoma is immunologically mediated: a comparison with squamous cell carcinoma. *Br J Dermatol.* 1994;131:789-798.
113. Melendez ND, Smoller BR, Morgan MVCAM. (CD-106) and ICAM (CD-54) adhesion molecules distinguish keratoacanthomas from cutaneous squamous cell carcinomas. *Mod Pathol.* 2003;16:8-13.
114. Tran TA, Ross JS, Boehm JR, Carlson JA. Comparison of mitotic cyclins and cyclin-dependent kinase expression in keratoacanthoma and squamous cell carcinoma. *J Cutan Pathol.* 1999;26:391-397.
115. Takeda H, Kondo S. Differences between squamous cell carcinoma and keratoacanthoma in angiotensin type-1 receptor expression. *Am J Pathol.* 2001;158:1633-1637.
116. Mukunyadzi P, Sanderson RD, Fan CY, Smoller BR. The level of syndecan-1 expression is a distinguishing feature in behavior between keratoacanthoma and invasive cutaneous squamous cell carcinoma. *Mod Pathol.* 2002;15:45-49.
117. Godbolt AM, Sullivan JJ, Weedon D. Keratoacanthoma with perineural invasion: a report of 40 cases. *Australas J Dermatol.* 2001;42:168-171.
118. Lapins NA, Helwig EB. Perineural invasion by keratoacanthoma. *Arch Dermatol.* 1980;116:791-793.
119. Basoglu Y, Metze D, Nashan D, Stander S. Keratoacanthoma with perineural invasion: an indicator for aggressive behavior? *J Dtsch Dermatol Ges.* 2008;6:952-955.
120. Abbas O, Hussein L, Kurban M, Kibbi AG. Plasmacytoid dendritic cell involvement in the host response against keratoacanthoma. *J Am Acad Dermatol.* 2014;70:1142-1145.
121. Zduniak K, Agrawal S, Symonowicz K, Jurczyszyn K, Ziolkowski P. The comparison of nuclear ubiquitous casein and cyclin-dependent kinases substrate (NUCKS) with Ki67 proliferation marker expression in common skin tumors. *Pol J Pathol.* 2014;65:48-54.
122. Slater M, Barden JA. Differentiating keratoacanthoma from squamous cell carcinoma by the use of apoptotic and cell adhesion markers. *Histopathology.* 2005;47:170-178.
123. Krunic AL, Garrod DR, Madani S, Buchanan MD, Clark RE. Immunohistochemical staining for desmogleins 1 and 2 in keratinocytic neoplasms with squamous phenotype: actinic keratosis, keratoacanthoma and squamous cell carcinoma of the skin. *Br J Cancer.* 1998;77:1275-1279.
124. Tran TA, Ross JS, Sheehan CE, Carlson JA. Comparison of oncostatin M expression in keratoacanthoma and squamous cell carcinoma. *Mod Pathol.* 2000;13:427-432.
125. Soddu S, Di Felice E, Cabras S, et al. IMP-3 expression in keratoacanthomas and squamous cell carcinomas of the skin: an immunohistochemical study. *Eur J Histochem.* 2013; 57:e6.
126. Klein-Szanto AJ, Barr RJ, Reiners JJ Jr, Mamrak MD. Filaggrin distribution in keratoacanthomas and squamous cell carcinoma. *Arch Pathol Lab Med.* 1984;108:888-890.
127. Ichikawa E, Ohnishi T, Watanabe S. Expression of keratin and involucrin in keratoacanthoma: an immunohistochemical aid to diagnosis. *J Dermatol Sci.* 2004;34:115-117.
128. LaPresto L, Cranmer L, Morrison L, Erickson CP, Curiel-Lewandrowski C. A novel therapeutic combination approach for treating multiple vemurafenib-induced keratoacanthomas: systemic acitretin and intralesional fluorouracil. *JAMA Dermatol.* 2013;149:279-281.
129. Reich A, Kobierzycka M, Wozniak Z, Cislo M, Szepietowski JC. Keratoacanthoma-like cutaneous metastasis of lung cancer: a learning point. *Acta Derm Venereol.* 2006;86:459-460.
130. Aramburu-Gonzalez JA, Rodriguez-Justo M, Jimenez-Reyes J, Santonja C. A case of soft tissue mesenchymal chondrosarcoma metastatic to skin, clinically mimicking keratoacanthoma. *Am J Dermatopathol.* 1999;21:392-394.
131. Cassarino DS, Xue W, Shannon KJ. Widespread cutaneous and perioral metastases of mesothelioma. *J Cutan Pathol.* 2003;30:582-585.
132. Lin JH, Lee JY. Primary cutaneous CD30 anaplastic large cell lymphoma with keratoacanthoma-like pseudocarcinomatous hyperplasia and marked eosinophilia and neutrophilia. *J Cutan Pathol.* 2004;31:458-461.
133. Kaddu S, Soyer HP, Cerroni L, Salmhofer W, Hodl S. Clinical and histopathologic spectrum of pilomatricomas in adults. *Int J Dermatol.* 1994;33:705-708.
134. Meffert JJ. Cutaneous sporotrichosis presenting as a keratoacanthoma. *Cutis.* 1998;62:37-39.
135. Lauermann F, Lyra M, Gaudio R. Sporotrichosis mimicking keratoacanthoma. *Am J Trop Med Hyg.* 2012;86:741.
136. Miteva L, Brostilova V, Schwartz RA. Verrucous systemic lupus erythematosus. *Acta Dermatovenerol Croat.* 2009;17: 301-304.
137. Misago N, Inoue T, Koba S, Narisawa Y. Keratoacanthoma and other types of squamous cell carcinoma with crateriform architecture: classification and identification. *J Dermatol.* 2013;40:443-452.
138. Kalilbian AE, Choos JN. Keratoacanthoma or verrucous carcinoma? A case report. *J Foot Ankle Surg.* 1993;32:584-590.
139. Shendrik I, Crowson AN, Magro CM. Follicular cutaneous squamous cell carcinoma: an under-recognized neoplasm arising from hair appendage structures. *Br J Dermatol.* 2013; 169:384-388.

140. Carr RA, Taibjee SM, Turnbull N, Attili S. Follicular squamous cell carcinoma is an under-recognized common skin tumor. *Diagn Histopathol.* 2014;20:289-296.
141. Griffiths RW. Keratoacanthoma observed. *Br J Plast Surg.* 2004;57:485-501.
142. Piscioli F, Boi S, Zumiani G, Cristofolini M. A gigantic, metastasizing keratoacanthoma. Report of a case and discussion on classification. *Am J Dermatopathol.* 1984;6: 123-129.
143. Davis BA, Monheit GD, Kline L. Metastatic skin cancer presenting as ptosis and diplopia. *Dermatol Surg.* 2006;32: 148-158.
144. Hodak E, Jones RE, Ackerman AB. Solitary keratoacanthoma is a squamous-cell carcinoma: three examples with metastases. *Am J Dermatopathol.* 1993;15:332-352.
145. Savage JA, Maize JC Sr. Keratoacanthoma clinical behavior: a systematic review. *Am J Dermatopathol.* 2014;36:422-429.
146. Weinstein MC, Brodell RT, Bordeaux J, Honda K. The art and science of surgical margins for the dermatopathologist. *Am J Dermatopathol.* 2012;34:737-745.
147. Rogers CR, Bentz ML. An evidence-based approach to the treatment of nonmelanoma facial skin malignancies. *Plast Reconstr Surg.* 2011;127:940-948.
148. Schell AE, Russell MA, Park SS. Suggested excisional margins for cutaneous malignant lesions based on Mohs micrographic surgery. *JAMA Facial Plast Surg.* 2013;15: 337-343.
149. Jackson JE, Kelly B, Petitt M, Uchida T, Wagner RF Jr. Predictive value of margins in diagnostic biopsies of nonmelanoma skin cancers. *J Am Acad Dermatol.* 2012;67:122-127.
150. Benest L, Kaplan RP, Salit R, Moy R. Keratoacanthoma centrifugum marginatum of the lower extremity treated with Mohs micrographic surgery. *J Am Acad Dermatol.* 1994;31:501-502.
151. Klein E, Helm F, Milgrom H, Stoll HL Jr, Traenkle HL. Tumors of the skin. II. Keratoacanthoma; local effect of 5-fluorouracil. *Skin.* 1962;1:153-156.
152. Kirby JS, Miller CJ. Intralesional chemotherapy for non-melanoma skin cancer: a practical review. *J Am Acad Dermatol.* 2010;63:689-702.
153. Annest NM, VanBeek MJ, Arpey CJ, Whitaker DC. Intralesional methotrexate treatment for keratoacanthoma tumors: a retrospective study and review of the literature. *J Am Acad Dermatol.* 2007;56:989-993.
154. Aubut N, Alain J, Claveau J. Intralesional methotrexate treatment for keratoacanthoma tumors: a retrospective case series. *J Cutan Med Surg.* 2012;16:212-217.
155. Martorell-Calatayud A, Requena C, Nagore E, et al. Intralesional infusion of methotrexate as neoadjuvant therapy improves the cosmetic and functional results of surgery to treat keratoacanthoma: results of a randomized trial [in Spanish]. *Actas Dermosifiliogr.* 2011;102:605-615.
156. Yeon JH, Jung JY, Choi JW, et al. Keratoacanthoma aggravated after photodynamic therapy. *J Dermatol.* 2010; 37:765-766.
157. Maydan E, Nootheti PK, Goldman MP. Development of a keratoacanthoma after topical photodynamic therapy with 5-aminolevulinic acid. *J Drugs Dermatol.* 2006;5:804-806.
158. Thiele JJ, Ziemer M, Fuchs S, Elsner P. Combined 5-fluorouracil and Er:YAG laser treatment in a case of recurrent giant keratoacanthoma of the lower leg. *Dermatol Surg.* 2004;30: 1556-1560.
159. Chaffai M, Houman MH, Haouet S, Ben Osman A. Keratoacanthoma centrifugum marginatum [in French]. *Ann Dermatol Venereol.* 1994;121:731-733.
160. V'Lckova-Laskoska MT, Laskoski DS. Keratoacanthoma centrifugum marginatum: a rare atypical variant of keratoacanthoma. *Clin Exp Dermatol.* 2008;33:259-261.
161. Bulj TK, Krunic AL, Cetner AS, Villano JL. Refractory aggressive keratoacanthoma centrifugum marginatum of the scalp controlled with the epidermal growth factor receptor inhibitor erlotinib. *Br J Dermatol.* 2010;163:633-637.
162. Farias MM, Hasson A, Navarrete C, Nicklas C, Garcia-Huidobro I, Gonzalez S. Efficacy of topical photodynamic therapy for keratoacanthomas: a case-series of four patients. *Indian J Dermatology Venereol Leprol.* 2012; 78:172-174.
163. Caccialanza M, Sopelana N. Radiation therapy of keratoacanthomas: results in 55 patients. *Int J Radiat Oncol Biol Phys.* 1989;16:475-477.
164. Cipollaro VA. The use of podophyllin in the treatment of keratoacanthoma. *Int J Dermatol.* 1983;22:436-440.
165. Starzycki Z. Use of Ufudix ointment for the treatment of keratoacanthoma. I. Clinical evaluation of the results [in Polish]. *Przegl Dermatol.* 1980;67:469-474.
166. Jeon HC, Choi M, Paik SH, Ahn CH, Park HS, Cho KH. Treatment of keratoacanthoma with 5% imiquimod cream and review of the previous report. *Ann Dermatol.* 2011;23: 357-361.
167. Kaźmierowski M, Bowszyc-Dmochowska M. Multiple keratoacanthomas treated with cryosurgery and retinoids. *Postepy Dermatol (Poznan).* 1997;14:397-402.
168. Street ML, White JW Jr, Gibson LE. Multiple keratoacanthomas treated with oral retinoids. *J Am Acad Dermatol.* 1990;23: 862-866.
169. Nofal A, Assaf M, Ghonemy S, Nofal E, Yosef A. Generalized eruptive keratoacanthoma: a diagnostic and therapeutic challenge. *Int J Dermatol.* 2015;54:160-167.
170. Reid DC, Guitart J, Agulnik M, Lacouture ME. Treatment of multiple keratoacanthomas with erlotinib. *Int J Clin Oncol.* 2010;15:413-415.
171. Sanders S, Busam KJ, Halpern AC, Nehal KS. Intralesional corticosteroid treatment of multiple eruptive keratoacanthomas: case report and review of a controversial therapy. *Dermatol Surg.* 2002;28:954-958.
172. Alloo A, Garibyan L, LeBoeuf N, et al. Photodynamic therapy for multiple eruptive keratoacanthomas associated with vemurafenib treatment for metastatic melanoma. *Arch Dermatol.* 2012;148:363-366.
173. Stevanović D, Krunic A. Keratoacanthoma: a dangerous trap. In: Panconesi E, ed. *Dermatology in Europe Proceedings of the First Congress of the European Academy of Dermatology and Venereology, Florence, Italy September 25-28, 1989.* Oxford: Blackwell Scientific; 1991:390-392.
174. Grossniklaus HE, Wojno TH, Yanoff M, Font RL. Invasive keratoacanthoma of the eyelid and ocular adnexa. *Ophthalmology.* 1996;103:937-941.
175. Schwartz RA. Keratoacanthoma: an abortive squamous cell carcinoma that does not always fail. *G Ital Dermatol Venereol.* 2003;138:355-362.